Novel *CRELD1* gene mutations in patients with atrioventricular septal defect

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**Background:** Atrioventricular septal defects (AVSDs) occur as clinical defects of several different syndromes, as autosomal dominant defects, and as sporadically occurring malformations. Consequently, there is genetic heterogeneity, but until recently, little is known about the genes involving in the pathogenesis of AVSD. *CRELD1* gene, a novel cell adhesion molecule, is a candidate gene for AVSD.

**Methods:** This study included 133 patients with AVSD and 200 healthy controls. Peripheral blood samples were collected and genomic DNA was extracted from the leukocytes. *CRELD1* was amplified by polymerase chain reaction (PCR) with specific primers. The sequences of PCR products were compared between the patients and controls.

**Results:** In a patient, a C-to-G transition was identified at nucleotide 857 in exon 8 that resulted in a substitution of alanine for proline at amino acid 286 in the first calcium-binding EGF domain. This patient had an isolated partial AVSD and the mutation was inherited from her mother. Another mutation was detected in a patient with a partial AVSD and evidence of Down syndrome. The heterozygous c.973G>A transition in exon 9 resulted in a substitution of lysine for glutamic acid at amino acid 325 (E325K) in the second calcium-binding EGF domain.

**Conclusions:** Two novel *CRELD1* mutations were identified in the calcium-binding EGF domain in patients with AVSD. *CRELD1* is likely to be an AVSD-susceptibility gene and *CRELD1* mutations may increase the risk of developing a heart defect rather than being a direct causative mutation.

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**Key words:** atrioventricular septal defect; *CRELD1*; Down syndrome

**Introduction**

Atrioventricular septal defect (AVSD), also known as atrioventricular canal defect or endocardial cushion defect, is a subset of malformation that constitutes 7%-8% of all recognized congenital heart diseases.[1] AVSD is sometimes referred to as an endocardial cushion defect because failure of endocardial cushion formation results in incomplete valvuloseptal morphogenesis. The atrioventricular endocardial cushion contributes to the formation of the inlet ventricular septum and atrial septum primum and the mitral and tricuspid valves. Consequently, genes involving in the development of the atrioventricular endocardial cushions have been recognized as prime candidates for AVSD.

To the present, two specific genetic loci for AVSD have been identified. The study of one large family led to the identification of the AVSD1 locus on chromosome 1p31-p21 (MIM 606215) by use of a combination of DNA pooling and shared segment analysis in a high-density genome screening.[2] Although the responsible gene has not yet been identified, this study demonstrated the existence of a congenital heart susceptibility gene inherited as an autosomal dominant trait with incomplete penetrance. In another study, analysis of chromosomal breakpoints in 3p-syndrome found the AVSD2 (MIM 606217) locus on chromosome 3p25.[3] The corresponding gene is *CRELD1* (GenBank accession number AF452623), which belongs to an epidermal growth factor-like family and encodes a cell surface protein that likely functions as cell adhesion molecule. *CRELD1* encodes a novel cell adhesion molecule that is expressed during cardiac cushion development.[4] To further investigate the role of *CRELD1* in AVSD
pathogenesis, we analyzed 133 patients with AVSD for CRELD1 mutations who had been admitted to Shanghai Children’s Medical Center from March 2007 to October 2009.

**Methods**

All participants (133 patients and 200 healthy controls) of this study were of the Chinese Han nationality. Blood samples were taken from the 133 patients aged from 2 months to 12 years. Clinical examination of the patients included anthropometric measurement, physical examination for dysmorphism and malformation, chest X-ray examination, electrocardiography, ultrasonic echocardiography, and some of the patients even underwent cardiac catheterization examination. The anatomic phenotypes of the patients are shown in Table 1. Among the 133 patients, 24 had Down syndrome, 2 had heterotaxy syndromes and polysplenia, 6 had heterotaxy syndromes and asplenia, and 101 had non-syndrome related AVSD. In 95 patients with complete AVSD, 35 were associated with other congenital heart abnormalities. Thirty-eight patients with partial AVSD were associated with clefting of the mitral valve. DNA samples from 200 unrelated and healthy individuals of the Chinese Han nationality (93 males and 107 females) served as controls. Before the study, informed consents were obtained from their parents or guardians of both patients and controls. The research protocols and procedures were approved by the Association of Medical Ethics of Shanghai Children’s Medical Center.

Peripheral blood samples were collected, and genomic DNA of each sample was extracted by use of Axygen DNA extraction kits (Axygen, Union City, CA). Primers were designed to amplify the entire CRELD1 coding region of 11 exons, including all intron-exon boundary junctions and at least 100 bp of intron covering potential splicing elements (Table 2). Polymerase chain reaction (PCR) products from each exon were tested and directly sequenced according to the ABI Big Dye Terminator Cycle Sequencing protocol and ABI 310 Automated Sequencer (ABI, Foster City, CA, USA).

BLAST search was used to identify homology between the sequences obtained from patients and to evaluate sequence conservation across species.

### Table 1. The anatomic phenotypes of the 133 patients included in the molecular screening

<table>
<thead>
<tr>
<th>Type of congenital heart defect</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>Non-DS complete AVSD</td>
<td>73</td>
</tr>
<tr>
<td>Isolated</td>
<td>41 (one with cleft, and one patient’s father with SAS)</td>
</tr>
<tr>
<td>With LSVC</td>
<td>2</td>
</tr>
<tr>
<td>With PDA</td>
<td>3</td>
</tr>
<tr>
<td>With DORV</td>
<td>5</td>
</tr>
<tr>
<td>With SV</td>
<td>9</td>
</tr>
<tr>
<td>With PA/VSD</td>
<td>5</td>
</tr>
<tr>
<td>With DORV and heterotaxy</td>
<td>8</td>
</tr>
<tr>
<td>Complete AVSD with DS</td>
<td>22</td>
</tr>
<tr>
<td>Isolated</td>
<td>19</td>
</tr>
<tr>
<td>With PDA</td>
<td>3</td>
</tr>
<tr>
<td>Non-DS partial AVSD</td>
<td>36</td>
</tr>
<tr>
<td>Isolated</td>
<td>35</td>
</tr>
<tr>
<td>With PDA</td>
<td>1</td>
</tr>
<tr>
<td>Partial AVSD with DS</td>
<td>2</td>
</tr>
<tr>
<td>Isolated</td>
<td>2</td>
</tr>
<tr>
<td>Total</td>
<td>133</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>SNP</th>
<th>Location</th>
<th>Aa change</th>
<th>Frequency in atrioventricular septal defect cases</th>
<th>Frequency in control subjects</th>
<th>rs number</th>
</tr>
</thead>
<tbody>
<tr>
<td>C1119T</td>
<td>Exon 10</td>
<td>H373H</td>
<td>39:133</td>
<td>41:200</td>
<td>rs 3774207</td>
</tr>
<tr>
<td>C383G</td>
<td>Exon 4</td>
<td>P128R</td>
<td>18:133</td>
<td>18:200</td>
<td>rs 2302787</td>
</tr>
</tbody>
</table>

**Table 2. Primers for the amplification of CRELD1 exons**

<table>
<thead>
<tr>
<th>Exons</th>
<th>Forward primer</th>
<th>Reverse primer</th>
<th>Amplicon (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>5′- CCTCTTTTGGCTTGTAATAAAGCAG</td>
<td>5′- CCACCTTAATCGCTGTGTTC</td>
<td>361</td>
</tr>
<tr>
<td>2</td>
<td>5′- GGGAAACACAGCGATTTAGTGGG</td>
<td>5′- ACCAGCAGAGACAGATTTGGC</td>
<td>341</td>
</tr>
<tr>
<td>3</td>
<td>5′- AGGAAAGGAGGGAGAGACTTGAG</td>
<td>5′- TGGGTTGGAGAAACGACTAC</td>
<td>318</td>
</tr>
<tr>
<td>4</td>
<td>5′- GGGAAATGGGAAACAGCCTATGTAC</td>
<td>5′- GCAAGCGGGATGGTCACAGAAG</td>
<td>297</td>
</tr>
<tr>
<td>5</td>
<td>5′- TGTGGTCTTCCACAGCCT</td>
<td>5′- TGGGAACACAGCGATTTAGTGGG</td>
<td>331</td>
</tr>
<tr>
<td>6</td>
<td>5′- TGGCCCTACTTTGAGGCGAGAC</td>
<td>5′- CCCATCCACACTCAGAGCAGAAG</td>
<td>468</td>
</tr>
<tr>
<td>7</td>
<td>5′- ATCCCGAGCGGACACATCC</td>
<td>5′- CAGCTCCTTCCACACCTCCA</td>
<td>226</td>
</tr>
<tr>
<td>8</td>
<td>5′- CTCGTCCTCCTCAAAAACCTT</td>
<td>5′- CAGCTCCTTCCACACCTCCA</td>
<td>331</td>
</tr>
<tr>
<td>9</td>
<td>5′- AGAGGGGAGTGTTGAGGAGATGGAC</td>
<td>5′- CTGATTGTGATGCGGAGTTC</td>
<td>446</td>
</tr>
<tr>
<td>10</td>
<td>5′- CGGGCTCTACATCGATCTCC</td>
<td>5′- AGGGTTAAAGGGGCTGGTGG</td>
<td>510</td>
</tr>
<tr>
<td>11</td>
<td>5′- TGTGGGAGGATTTGCGGGGAGAC</td>
<td>5′- TACCTGGGCTGCTGTGAAGG</td>
<td>441</td>
</tr>
</tbody>
</table>
Results
A heterozygous c.857C>G mutation was identified in one patient with isolated partial AVSD (Fig. 1). The mutation resulted in a substitution of arginine for proline at amino acid 286 (P286R) in the first calcium-binding EGF domain. After exclusion in 200 normal chromosomes of the Chinese population we considered the variant to be a novel CRELD1 mutation in patients with AVSD. Analysis of genomic DNA from the parents of the proband showed that this mutation was inherited from the mother. Two-dimensional echocardiography and color flow Doppler studies showed that the mother had structurally normal heart with normal ventricular function, demonstrating incomplete penetrance of this CRELD1 variant.

The second mutation was identified in a patient with Down syndrome, a partial AVSD and pulmonary hypertension. The heterozygous c.973G>A transition in exon 9 resulted in a substitution of lysine for glutamic acid at amino acid 325 (E325K) in the second calcium-binding EGF domain (Fig. 2). The parents of the proband refused to do genetic analysis. The variant was not detected in 200 normal chromosomes of the Chinese population.

Cross-species analysis demonstrated that these nucleotides and codons were conserved: >90% homology in regions of the CRELD1 gene (Fig. 3). In addition, we detected two previously reported polymorphisms (rs 2302787 and rs 3774207) (Table 3).

Discussion
AVSDs include a spectrum of anomalies of the atrioventricular valves and the atrial and ventricular septa. In the complete form there are a single common atrioventricular valve, an atrial septal defect (ostium primum), and a confluent posterior ventricular septal defect in the inlet portion of the ventricular septum. Without intervention, few patients survive to 5 years of age due to the early onset of heart failure and irreversible pulmonary hypertension. In the partial form there are two separate right and left atrioventricular valves with a "cleft" of the mitral valve, an atrial septal defect (ostium primum), and no ventricular septal communication.

From the pathogenetic point of view, this malformation is classified in a group of defects of the extracellular matrix. The atrioventricular septal defect is a "classic" heart defect in children with Down syndrome and is one of the congenital heart diseases frequently associated with extracardiac anomalies. The evident genetic basis for this malformation is demonstrated also by the prevalence of patients with heterotaxy, other chromosomal defects or Mendelian syndromes. Based on anatomical, clinical, and genetic observations, four major groups of patients with atrioventricular canal defect have been recognized: group I, Down syndrome (45%); group II, heterotaxy syndrome (15%); group III, other syndromes with situs solitus (15%), such as 3p-syndrome, CHARGE syndrome; and group IV,
nonsyndromic patients (25%). Different genetic bases could explain the anatomical differences in the various genetic groups of patients with AVSD. Several families have been found to have autosomal-dominant AVSD with incomplete penetrance,\(^{[15,16]}\) demonstrating that AVSD can be inherited as a single gene defect. But the majority cases (about 65%) of AVSD is sporadic.\(^{[11]}\) The relatively high incidence of sporadic cases indicates that AVSD is usually genetically complex, with multiple factors contributing to susceptibility and variability of expression.

The CRELD1 family consists of two matricellular proteins thought to be involved in cell adhesion processes. Identification and characterization of the CRELD1 gene on chromosome 3p25 led to the establishment of CRELD1 as the first known genetic risk factor for isolated AVSD.

In 52 patients with non-Down syndrome-related AVSD, Robinson et al.\(^{[17]}\) identified 3 CRELD1 mutations (p. R107H, p.T311I, p.R329C) in those patients with partial AVSD. Zatyka et al.\(^{[18]}\) identified another missense mutation (p. P162A) in an individual with partial AVSD. Sarkozy et al.\(^{[19]}\) found no pathogenic mutations in 31 individuals with isolated AVSD. But Posch et al.\(^{[20]}\) did not identify any mutations in 16 AVSD patients. Our studies and previous reports of CRELD1 gene screening showed that the incidence of CRELD1 gene mutations in non-Down syndrome-related AVSD is 2.0% (5/253 patients analyzed). In the non-Down syndrome group, CRELD1 mutations were detected exclusively in patients with partial AVSD (5/121; 4.1%; not including Posch’s cases), suggesting a genotype-phenotype correlation with this specific anatomic AVSD subtype. In our study, the proband inherited the CRELD1 P286R mutation from the mother who had a structurally and functionally normal heart. As reported previously, these findings are consistent with incomplete penetrance.\(^{[21]}\) The presence of unaffected family members carrying a CRELD1 mutation showed that CRELD1 mutation is neither necessary nor sufficient to cause AVSD, indicating that mutation of CRELD1 increases susceptibility for AVSD rather than serves as a monogenic cause.

AVSD is commonly associated with Down syndrome. Among children with a normal karyotype, the frequency of AVSD is 1 in 10 000 live births, but is 2000 in 10 000 live births in the Down syndrome population.\(^{[22]}\) The study of the relationship between CRELD1 mutation and the occurrence of AVSD on the genetic background of trisomy 21 was rare. Maslen et al.\(^{[21]}\) identified 2 heterozygous missense mutations (R329C, E414K) in 2 subjects with Down syndrome and AVSD. One had partial AVSD with a secundum atrial septal defect, patent ductus arteriosus, tricuspid regurgitation and pulmonary hypertension; the other had complete AVSD and tricuspid regurgitation. Interestingly, with the same R329C mutation, the severity of the heart defect was greater in a patient with Down syndrome, who had a complete AVSD and pulmonary hypertension, compared to the patient with an isolated partial AVSD. These data and ours indicate that the incidence of CRELD1 mutation in patients with AVSD associated with Down syndrome is 4.8% (3/63). There is a significant difference in the frequency of CRELD1 mutation between patients with Down syndrome and those without Down syndrome (\(\chi^2=1.586, P=0.208\)). The results indicate that the mutation of CRELD1 gene may contribute to the pathogenesis of AVSD in the context of trisomy 21, possibly through genetic interaction with undetermined causative gene(s) on chromosome 21 and environmental factors.

The two novel mutations associated with AVSD occurred in calcium-binding EGF domains. Up to now, 7 CRELD1 mutation sites associated with AVSD have been identified and 4 of them are in calcium-binding EGF domains (Fig. 4). The calcium-binding EGF domain is prevalent in many extracellular proteins, and heterozygous mutations affecting calcium-binding EGF domains have been associated with a number of genetic disorders. Calcium-binding EGF domains are highly conserved with specific disulfide bonding patterns. The lysine-to-arginine and glutamic acid-to-proline substitutions in calcium-binding EGF domains, the change of the amino acid polarity, would be expected to interfere with protein folding and further affect the protein function.

In conclusion, the present mutation analysis of CRELD1 gene in 133 Chinese AVSD patients revealed 2 novel mutations in the calcium-binding domain. To our knowledge, the sample size in our study is the largest. The overall mutation rate was low (2/133, 1.5%) and unaffected family members carrying a CRELD1 mutation indicate that CRELD1 is an AVSD-
susceptibility gene and that CRELD1 mutations increase the risk of developing a heart defect rather than serve as direct causative mutations.

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Ethical approval: This protocol was approved by the Hospital Ethical Committee.

Competing interest: None.

Contributors: Guo Y wrote the main body of the article. Sun K and Shen J are supervisors of this article. Guo Y and Yuan L contributed equally to this paper. Li F and Wang J provided advice on medical aspects.

References

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